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**Effect of Boiling Time on Content of The Total Flavonoid of Kitolod (*Isotoma  
longiflora* (L.) C. Presl.)**

**Penulis:**

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**PROGRAM STUDI ADMINISTRASI KESEHATAN  
FAKULTAS KESEHATAN DAN TEKNOLOGI  
UNIVERSITAS MUHAMMADIYAH KLATEN**

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Dear Authors: Muchson Arrosyid\*, Choiril Hana Mustofa, Sutaryono, and Alifiasri Praptiwi Rohmah

We are pleased to inform you that your abstract (ABS-184, Oral Presentation), entitled:

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has been reviewed and accepted to be presented at ICoSHEET 2019 conference to be held on 18-19  
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# THE EFFECT OF BOILING TIME ON TOTAL FLAVONOID LEVELS OF KITOLOD LEAF (*Isotoma longiflora* (L.) C. Presl.)

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# INTRODUCTIONS

Kitolod leafes can be used as a healing medicine for wounds, asthma, bronchitis, rheumatism and so on. Boiling is usually done by the community to increase the acceptance of food and traditional medicines.

Kitolod leafes containing chemical compounds : phenolic alkaloids, terpenoids, steroids and flavonoids.

Boiling on traditional medicinal ingredients usually has a destructive effect on flavonoids and phenolics compounds because both are unstable components. It is suspected that boiling can affect the levels of flavonoids in kitolod leafes.

From the above statement, it is necessary to study the effect of boiling time on flavonoid levels of kitolod leafes due to boiling heat. The research treatment with a variation of boiling time is 5 minutes, 10 minutes, 15 minutes and 20 minutes.

# Research methods

The research treatment with a variation of boiling time is 5 minutes, 10 minutes, 15 minutes and 20 minutes.

Quantitative analysis test for flavonoid content of kitolod leafes used Spectrophotometry method, with UV-Vis spectrophotometer instrument. ShiMadzu UV-1800.

# Observation Data

Determination of total flavonoid levels by absorption at a maximum wavelength of 431 nm obtained total flavonoid levels:

<b>Boiling Time</b>	<b>Total Flavanoid Levels</b>
5 minutes :	0.3199% w / w or 31.9 mg / kg,
10 minutes :	0.2563% w / w or 25.6 mg / kg,
15 minutes :	0.2239% w / w or 22.3 mg / kg,
20 minutes :	0.1725% w / w or 17.2 mg / kg

# Data analysis

Based on observational data obtained were tested statistically using the One-Kolmogorov-Smirnov Test obtained significance  $0.992 > 0.05$  ( $H_0$  accepted).

Then proceed with the One Way Anova test, the homogeneity value is obtained that is  $0.504 > 0.05$  ( $H_0$  is accepted).

Anova test data shows that the significance of  $0.000 < 0.05$  then  $H_0$  is rejected, followed by the LSD test.

From the data between treatments it can be said that there is a significant difference between treatments, namely at each boiling time comparison there is a decrease in flavonoid levels.

# Conclusion

- ▶ The length of boiling time affects the total flavonoids levels of kitolod leafes (*Isotoma longiflora* (L.) C. Presl), the longer the leafes are boiled, the compound content of flavonoid levels decreases.



## Panduan Revisi Article ICoSHEET.

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# Effect of EFFECT OF BOILING Time ON On CONTENT Content Off THE The TOTAL Total FLAVONOID Flavonoid OF Of KITOLOD Kitolod (*Isotoma longiflora* (L.) C. Presl.)

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## AbstractBSTRACT

Kitolod (*Isotoma longiflora* (L.) C. Persl) is a annual herbaceous plant that usually grows on the edge of a moist wall. The leaves of the kitolod plant can be used as medicine for healing wounds, asthma, bronchitis, rheumatism and so on. Kitolod contains chemical compounds namely alkaloids namely lobelamin, isotomine and lobelin. In the leaf part of kitolod contains chemical compounds namely alkaloids, saponins, polyphenols and flavonoids.

This study aims to determine the effect of boiling time on the total flavonoid content of water extract of kitolod leaves. Extraction was done by boiling, the determination of the total amount of flavonoids was carried out by the AlCl<sub>3</sub> method and then the absorption was measured using Spectrophotometry UV-Vis with a wavelength of 431 nm.

The results showed that qualitatively the samples contained flavonoids by showing red. Quantitatively the kitolod leaves are boiled with longer time, the total flavonoid content will decrease. The total flavonoid content of kitolod leaves in boiling 5 minutes obtained an average level of 0.31995% w/w or 31.9 mg/kg, boiling 10 minutes 0.25635% w/w or 25.6 mg/kg, boiling 15 minutes 0.2239% w/w or 22.3 mg/kg, boiling 20 minutes 0.17254% w/w or 17.2 mg/kg.

**Keywords:** kitolod leaves, boiling, flavonoid content, Spectrophotometry UV-Vis

## 1. INTRODUCTIONntroduction

Medicinal plants and traditional medicines are an asset that needs to be constantly explored, one of the plants that has the potential to become a medicinal plant. The leaves of the kitolod plant can be used as a remedy for wound healing, asthma, bronchitis, rheumatism and so on (Safitri *et al.*, 2009)[6]. Kitolod plants (*Isotoma longiflora* (L.) C. Persl) contain chemical compounds including alkaloid

compounds namely lobelamin, isotomin and lobelin. Kitodod plants contain other metabolite compounds such as phenol, terpenoids and steroids [(Paramita *et al.*, 20154)]. In addition, kitolod plants also contain flavonoid compounds (Utami *et al.*, 2013)[8].

Regular reductions are made by the community to increase acceptance of traditional food and medicine. Heat processes such as decomposition cause changes in the structural and cellular matrix interactions that have a negative and positive effect on the phytochemical content. The heat treatment of food and tracers drugs usually has a destructive effect on flavonoid and phenolic compounds as they are both unstable components. It is thought that boiling can affect the levels of flavonoids in the leaves of the kitolod (Saika and Mahanta, 2013)[7]

Flavonoids are secondary metabolite compounds that are formed through the ammonia pathway. This compound is manufactured from sinnaoil-CoA units by extension the chain using 3 malonil-CoA. The khalkhon synthase enzyme combines this compound into the khalkon. Khalkon is a precursor of flavonoid derivatives in many plants. (Dewick, 2002).

Flavonoid levels can be set by different methods, each analytical method having different levels of advantage. One of the methods used is the UV-Vis Spectrophotometry method. Some of the factors underlying this selection are speed, accuracy, sensitivity, accuracy, selectivity and practicality. The UV-Vis spectrophotometry method is used to determine concentration of the compounds. UV-Vis spectrophotometers are instruments that combine wavelengths and frequencies of absorption intensity (transmission or absorbance) and are expressed in the form of a spectrum or absorption band. The formation of absorption bands is due to the excitation of more than one type of electron in a very complex group of molecules (Gandjar and Rohman, 2010).

[2].

## 2. RESEARCH Research METHODSMethods

### 2.1. Tools and Materials

Tools used in this study include a set of glass appliances (Pyrex), UV-Vis Spectrophotometer 1800 (Shimadzu), electric scales (Ohaus), Quartz quartz, vortex, incubators, mixing sticks, electric stoves. Materials used in this study were: Kitolod leaves (*Isotoma longiflora* (L.) C. Presl.) Dried to 200 grams, technical methanol solvents, methanol *pa*, ethanol *pa*, quercitinquercetine, aquadest, AlCl<sub>3</sub> 10% solution, 1 M potassium acetate, HCl concentrated, Mg.

### 2.2. Determined plantCrop

Determination is made to determine the identity of the plant material used in the research, namely kitolod leaves obtained from the villagers located in East Talang, Talang Village, Bayat District, Klaten District.

### 2.3. Powder simplification simplisia making

Fresh kitolod leaves of 4000 g were washed with running water and then rinsed, then dried under sunlight covered with black cloth. Dried kitolod leaves are reduced in size (Fazil *et al.*, 2017[1])

### 2.4. Extract Making

The production of Kkitolod leaves water extract is made by means of boiling with varying degrees of boiling time. 50 grams of simplified powdered kitolod leaves were added to 250 mL boiled boil until boiling time was 5 minutes, 10 minutes, 15 minutes and 20 minutes while stirring occasionally. The infusion results are filtered hot and squeezed (Puspitasari and Prayogo, 2016).[5]

The extracts obtained are calculated by means of:

Randemen ending: (weight of the extract) / (weight of simplificationsimplisia) x 100%

### 2.5. Characterization of organoleptic extract characteristics

The organoleptic parameters of the extract are descriptive, color, taste and smell (Anonymous, 2000).

### 2.6. Flavonoid Qualitative Test

The extracts were extracted 1 ml in a reaction tube and heated on a flame of flame, and then dissolved in 1 ml of 70% ethanol, followed by a small amount of Mg metal and 3–5 drops of concentrated HCl. Positive results when formed in red, orange or yellow solution indicate the presence of flavonoids. (Halimah, 2010).

## 2.7. Flavonoid Quantitative Test

### a.2.7.1. Determination of Maximum Wave Length

10,0 mg quercetin was dissolved in 100 ml of methanol (1000 ppm concentration) as a stock solution. 1 ml of quercetin solution (quartz) was diluted with 2 ml of methanol and then 10% 1 mL aluminum (III) chloride was added. After waiting for 30 minutes, the absorbance of the comparison solution was measured with a UV-visible spectrophotometer at 380 nm – 780 nm. Created a calibration curve and obtained linear regression (Gandjar and Rohman, 2007[2])

### 2.7.2.b. Operating time

The assay was performed by mixing 1.0 ml of quenching quercetine solution with methanol to a volume of 5.0 ml in a squash, the mixture was shaken and kept for 5 minutes. The solution was measured at a 431.0 nm wavelength with a 45 minute stabilization time (Windasari and Ari, 2013[9]).

### 2.7.3.c. The production of a standard curve of quercetine quenching

A total of 10 mg of quenching quercetin was dissolved in 100 ml of methanol as a stock solution. The solution quenching of the quench quercetine was made at a concentration of 10, 20, 30, 40, 50 ppm. A total of 1 ml of quenching quercetine solution of each concentration was added with 1 ml of aluminum (III) chloride 10%, 1 ml Potassium acetate 1 M and added aquadest to 10 ml. For 15 minutes, the absorbance reader was readed absorbed using a UV-Vis Spectrophotometer at 431 nm (Gandjar and Rohman, 2007)

[2]

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### 2.7.4d. Total Flavonoid quantitative assay test

A total of 5,0 grams of the extract sample was dissolved in methanol at a concentration of 500 ppm. 1 ml of the extracts were added with 2 ml of methanol, then 1 ml of 10% aluminum (III) chloride, 1 ml of 1 M sodium acetate, and aquadest volume to 10 ml in volume. After incubation for 30 minutes, the absorbance of the comparison solution was measured with a spectrophotometer at the measured wavelengths (431,0 nm). It is then calculated using linear regression equations of the quadratic quercetine calibration curve which has also been measured previously (Ganjar and Rohman, 2007[2]).

$$y = b x + a$$

Description:

y = absorbance

a = intersep

b = slope

x = concentration

This assay was used to determine the levels of flavonoids in the kitolod leaves (*Isotoma longiflora* (L) C. Persl).

$$K = (C.V.FP) / BS$$

Results obtained are converted to%

K = Flavonoid rate level (%)

C = Concentration (ppm)

V = Sample volume (ml)

Fp = Dilution factor  
BS = Sample weight (grams)

### 2.8. Data Analysis

Concentration series data made from standard quenching quercetine are then made by the standard curve equation. The standard curve equation  $y = bx + a$  with the description  $y =$  absorbance,  $x =$  the rate in units of ppm (mg / ml). Absorbance of the obtained leaves extract extract was included in the standard curve equation to obtain the total flavonoid content of the kitolod leaves.

Statistical analyzes were performed using ANOVAAnova, to determine dispersal normality and homogeneity were analyzed using Kolmogrov-Smirnov test. The obtained data are normal and uniform when the values are  $P > 0.05$ . To find out whether there is a significant difference or not, then LSD analysis is useful to determine the difference between the average difference between treatments.

## 3. RESULTS Results AND and DISCUSSIONDiscussion

### 3.1. Determined plant

The results of the plant determination indicate that this plant belongs to the family Lobeliaceae, a species (*Isotoma longiflora* (L.) C. Presel)

### 3.2. Extract Results

The extraction results of kitolod leaves (*Isotoma longiflora* (L.) C. Presel) were performed by varying the 5-minute, 10-minute, 15-minute, and 20-minute sterilization boiling method with aquadest solvent. 50 grams of dried kitolod leaves were obtained from 13.9 grams, 12.2 grams, 12.1 grams, 14.3 grams. The leaves extract of kitolod leaves (*Isotoma longiflora* (L.) C. Presel) obtained in dark brown and has an aromatic, bitter taste.

Table 1. Rendering Randemen Results

Treatment	Weight sample	Weight Extract (g)	Randemen (%)
5 minutes	50	13.9	27.8 %
10 minutes	50	12.2	24.4 %
15 minutes	50	12.1	24.2 %
20 minutes	50	14.3	28.6 %

Data source: Primary Data, 2019

### 3.3. Flavor Flavonoid qualitative test results

Qualitative identification results of flavonoid leaf flavonoids containing flavonoids by showing that leaf extract extract (*Isotoma longiflora* (L.) C. Presel) reacted with yellow solid Mg and HCl powder. This is in line with the study conducted by Hapsari Atika (2016) where at the time of addition of reagents appeared yellow indicating the presence of flavonoid compounds. This is because flavonoids include phenolic compounds, when the phenol is treated with the base it forms a color due to the conjugation system of the aromatic group.

Flavor qualitative test results

Qualitative flavonoids identification of results of flavonoid kitolod leaves (*Isotoma longiflora* (L.) C. Presel) flavonoids containing flavonoids positively by showing a yellow formed, when extract that leaf extract extract (*Isotoma longiflora* (L.) C. Presel) reacted with powder yellow solid Mg and HCl concentrated powder. This is in line with the study conducted by Hapsari Atika (2016)[3] where at the time of addition of reagents appeared yellow indicating the presence of flavonoid compounds. This is because flavonoids include phenolic compounds, when the phenol is treated with the base it forms a color due to the conjugation system of the aromatic group.

### 3.4. Maximum Wave Length and Absorption

Table 2. Maximum Wave Length and Absorption

Absorbance wavelength (nm)	Absorbance
431	0.7450

Absorbance wavelength (nm)

431 0.7450

Data source: Primary Data, 2019

Setting the maximum wavelength for measurement at the maximum absorption wavelength will result in maximum absorption, at the maximum wavelength having maximum sensitivity due to the greatest absorbance change. The resultant maximum wavelength of the standard solution was 431.0 nm with an absorbance of 0.7450.

### 3.5. Operating Time (OT) setting

Operating time was performed with the aim of knowing the stable measurement time when the sample reacted perfectly to the color reagent, the stabilization time being 45 minutes.

Table 3 Operating Time (OT)

Time (minute)	Absorbance
10	0.782
11	0.782
12	0.782
13	0.781
14	0.781
15	0.781
16	0.781
17	0.781
18	0.781
19	0.780
20	0.781
21	0.780
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23	0.781
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36	0.779
37	0.779
38	0.779
39	0.779
40	0.779
41	0.779
42	0.779

3.6.

*Standard Curve Making*

The results of the concentration and absorbance of the standard curve solution can be seen in table 43:

Table 43. Concentration of Standard Curve Solution

<u>Concentration (ppm)</u>	<u>Absorbance</u>
10	0.259
20	0.330
30	0.403
40	0.485
50	0.562

Concentration (ppm) Absorbance

10 0.259

20 0.330

30 0.403

40 0.485

50 0.562

Data source: 2019 Primary Data

The concentration variation is used to detect differences in absorbance, where the higher the concentration of absorbance values the greater.

The production of a standard curve aims to obtain the standard solution of equilibrium in the determination of the sample rate, the relationship between the quenching quercetin concentration and the absorption, the data obtained a correlation coefficient (r) of 0.9993 approaching figure 1 shows that the regression equation is linear.

Table 5. Standard Curves

Data Source: Primary Data 2019

Based on table 5 43 we obtain  $y = 0.0076x + 0.1795$  with values of  $r = 0.9993$  indicating that these values are linear.

### 3.7. Determination of Flavonoid RatesLevel

Determination of total flavonoid readings with absorbance at a maximum wavelength of 431 nm was obtained with total flavonoid content of kitolod leaves (*Isotoma longiflora* (L.) C. Presl) of 5 min incubation at an average rate of 0.31995% b w / w b or 31.9 mg / kg, 10 min infusion 0.25635% b w / b w or 25.6 mg / kg, 15 min 0.2239% b w / bw or 22.3 mg / kg, 20 min 0.17254% b w / bw or 17.2 mg / kg (table 6). Based on the observational data obtained test statistically using the One-Kolmogorov-Smirnov Test obtained  $0.992 > 0.05$  ( $H_0$  accepted). Then, with the one-way Anova test, homogeneity was found to be  $0.504 > 0.05$  ( $H_0$  accepted). From the Anova test data show that the sign of  $0.000 < 0.05$  then  $H_0$  is rejected, followed by LSD test. From the data between treatments it can be said that there is a significant difference between the treatments that is in each comparison of the time of reduction of flavonoid levels. (table 7)

## 4. CONCLUSIONConclusion

The longer the fertilization boiling time affects the total flavonoid content of the leaves kitolodf (*Isotoma longiflora* (L.) C. Presl), the longer the decomposition of the flavonoid content compound decreases.

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29 Januari 2020

Ke  
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Perihal: Penyampaian revisi artikel

Kami mengucapkan banyak terima kasih kepada ICoSHEET 2019 Organizing Committee atas kesempatan untuk mengirimkan kembali revisi naskah kami yang berjudul Effect of Boiling Time on Content of The Total Flavonoid of Kitolod (*Isotoma longiflora* (L.) C. Presl.)

Sebagai pertimbangan dan informasi bagi reviewer, naskah ini memuat semua masukan dan rekomendasi reviewer untuk menyempurnakan artikel.

Terima kasih atas diterimanya naskah revisi kami.

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## Lampiran Revisi Outhor untuk Reviewer

### Effect of Boiling Time On Content Of The Total Flavonoid Of Kitolod (*Isotoma longiflora* (L.) C. Presl.)

Muchson Arrosyid<sup>1)</sup>, Choiril Hana Mustofa<sup>1)</sup>, Sutaryono<sup>1)</sup> Alifiasri Praptiwi Rohmah<sup>1)</sup>

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#### Abstract

Kitolod (*Isotoma longiflora* (L.) C. Persl) is a annual herbaceous plant that usually grows on the edge of a moist wall. The leaves of the kitolod plant can be used as medicine for healing wounds, asthma, bronchitis, rheumatism and so on. Kitolod contains chemical compounds namely alkaloids namely lobelamin, isotomine and lobelin. In the leaf part of kitolod contains chemical compounds namely alkaloids, saponins, polyphenols and flavonoids.

This study aims to determine the effect of boiling time on the total flavonoid content of water extract of kitolod leaves. Extraction was done by boiling, the determination of the total amount of flavonoids was carried out by the AlCl<sub>3</sub> method and then the absorption was measured using Spectrophotometry UV-Vis with a wavelength of 431 nm.

The results showed that qualitatively the samples contained flavonoids by showing red. Quantitatively the kitolod leaves are boiled with longer time, the total flavonoid content will decrease. The total flavonoid content of kitolod leaves in boiling 5 minutes obtained an average level of 0.31995% w/w or 31.9 mg/kg, boiling 10 minutes 0.25635% w/w or 25.6 mg/kg, boiling 15 minutes 0.2239% w/w or 22.3 mg/kg, boiling 20 minutes 0.17254% w/w or 17.2 mg/kg.

**Keywords:** kitolod leaves, boiling, flavonoid content, Spectrophotometry UV-Vis

#### 1. Introduction

Medicinal plants and traditional medicines are an asset that needs to be constantly explored, one of the plants that has the potential to become a medicinal plant. The leaves of the kitolod plant can be used as a remedy for wound healing, asthma, bronchitis, rheumatism and so on[6]. Kitolod plants (*Isotoma longiflora* (L.) C. Persl) contain chemical compounds including alkaloid compounds namely lobelamin, isotomin and lobelin. Kitodod plants contain other metabolite compounds such as phenol, terpenoids and steroids [4]. In addition, kitolod plants also contain flavonoid compounds [8].

Regular reductions are made by the community to increase acceptance of traditional food and medicine. Heat processes such as decomposition cause changes in the structural and cellular matrix interactions that have a negative and positive effect on the phytochemical content. The heat treatment of food and tracteric drugs usually has a destructive effect on flavonoid and phenolic compounds as they are both unstable components. It is thought that boiling can affect the levels of flavonoids in the leaves of the kitolod [7]

Flavonoids are secondary metabolite compounds that are formed through the ammonia pathway. This compound is manufactured from sinaldoil-CoA units by extension the chain using 3 malonil-CoA. The khalkhon synthase enzyme combines this compound into the khalkhon. Khalkhon is a precursor of flavonoid derivatives in many plants.

Flavonoid levels can be set by different methods, each analytical method having different levels of advantage. One of the methods used is the UV-Vis Spectrophotometry method. Some of the factors underlying this selection are speed, accuracy, sensitivity, accuracy, selectivity and practicality. The UV-Vis spectrophotometry method is used to determine concentration of the compounds. UV-Vis spectrophotometers are instruments that combine wavelengths and frequencies of absorption intensity (transmission or absorbance) and are expressed in the form of a spectrum or absorption band. The formation of absorption bands is due to the excitation of more than one type of electron in a very complex group of molecules [2].

## 2. Research Methods

### 2.1. Tools and Materials

Tools used in this study include a set of glass appliances (Pyrex), UV-Vis Spectrophotometer 1800 (Shimadzu), electric scales (Ohaus), quartz, vortex, incubators, mixing sticks, electric stoves.

Materials used in this study were: Kitolod leaves (*Isotoma longiflora* (L.) C. Presl.) Dried to 200 grams, technical methanol solvents, methanol *pa*, ethanol *pa*, quercetine, aquadest, AlCl<sub>3</sub> 10% solution, 1 M potassium acetate, HCl concentrated, Mg.

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### 2.2. Determined *plant*

Determination is made to determine the identity of the plant material used in the research, namely kitolod leaves obtained from the villagers located in East Talang, Talang Village, Bayat District, Klaten District.

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### 2.3. Powder *simplisia* making

Fresh kitolod leaves of 4000 g were washed with running water and then rinsed, then dried under sunlight covered with black cloth. Dried kitolod leaves are reduced in [1]

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Kitolod leaves water extract is made by means of boiling with varying degrees of boiling time. 50 grams of simplified powdered kitolod leaves were added to 250 mL boiled boil until boiling time was 5 minutes, 10 minutes, 15 minutes and 20 minutes while stirring occasionally. The infusion results are filtered hot and squeezed [5]

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$\text{Randemen} = (\text{weight of the extract}) / (\text{weight of simplisia}) \times 100\%$

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### 2.5. Characterization of organoleptic extract characteristics

The organoleptic parameters of the extract are descriptive, color, taste and smell.

### 2.6. Flavonoid Qualitative Test

The extracts were extracted 1 ml in a reaction tube and heated on a flame of flame, and then dissolved in 1 ml of 70% ethanol, followed by a small amount of Mg metal and 3–5 drops of concentrated HCl. Positive results when formed in red, orange or yellow solution indicate the presence of flavonoids.

### 2.7. Flavonoid Quantitative Test

#### 2.7.1. Determination of Maximum Wave Length

10,0 mg quercetin was dissolved in 100 ml of methanol (1000 ppm concentration) as a stock solution. 1 ml of quercetin solution (quartz) was diluted with 2 ml of methanol and then 10% 1 mL aluminum (III) chloride was added. After waiting for 30 minutes, the absorbance of the comparison solution was measured with a UV-visible spectrophotometer at 380 nm – 780 nm. Created a calibration curve and obtained linear regression [2]

### 2.7.2. Operating time

The assay was performed by mixing 1.0 ml of quercetine solution with methanol to a volume of 5.0 ml in a squash, the mixture was shaken and kept for 5 minutes. The solution was measured at a 431.0 nm wavelength with a 45 minute stabilization time [9].

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A total of 10 mg of quercetin was dissolved in 100 ml of methanol as a stock solution. The solution of the quercetine was made at a concentration of 10, 20, 30, 40, 50 ppm. A total of 1 ml of querecetin solution of each concentration was added with 1 ml of aluminum (III) chloride 10%, 1 ml Potassium acetate 1 M and added aquadest to 10 ml. For 15 minutes, the absorbance was readed using a UV-Vis Spectrophotometer at 431 nm [2]

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### 2.7.4. Total Flavonoid quantitative test

A total of 5.0 grams of the extract sample was dissolved in methanol at a concentration of 500 ppm. 1 ml of the extracts were added with 2 ml of methanol, then 1 ml of 10% aluminum (III) chloride, 1 ml of 1 M sodium acetate, and aquadest to 10 ml in volume. After incubation for 30 minutes, the absorbance of the comparison solution was measured with a spectrophotometer at the measured wavelengths (431,0 nm). It is then calculated using linear regression equations of the quercetine calibration curve which has also been measured previously [2].

$$y = b x + a$$

Description:

y = absorbance

a = intersep

b = slope

x = concentration

This assay was used to determine the levels of flavonoids in the kitolod leaves (*Isotoma longiflora* (L) C. Persl).

$$K = (C.V.FP) / BS$$

Results obtained are converted to%

K = Flavonoid level (%)

C = Concentration (ppm)

V = Sample volume (ml)

Fp = Dilution factor

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### 2.8. Data Analysis

Concentration series data made from standard quercetine are then made by the standard curve equation. The standard curve equation  $y = bx + a$  with the description y = absorbance, x = the rate in units of ppm (mg / ml). Absorbance of the obtained leaves extract was included in the standard curve equation to obtain the total flavonoid content of the kitolod leaves.

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Statistical analyzes were performed using Anova, to determine dispersal normality and homogeneity were analyzed using Kolmogrov-Smirnov test. The obtained data are normal and uniform when the values are  $P > 0.05$ . To find out whether there is a significant difference or not, then LSD analysis is useful to determine the difference between the average difference between treatments.

## 3. Results and Discussion

### 3.1. Determined plant

The results of the plant determination indicate that this plant belongs to the family Lobeliaceae, a species (*Isotoma longiflora* (L.) C. Presel)

### 3.2. Extract Results

The extraction results of kitolod leaves (*Isotoma longiflora* (L.) C. Presel) were performed by varying the 5-minute, 10-minute, 15-minute, and 20-minute boiling method with aquadest solvent. 50 grams of dried kitolod leaves were obtained from 13.9 grams, 12.2 grams, 12.1 grams, 14.3 grams. The extract of kitolod leaves (*Isotoma longiflora* (L.) C. Presel) obtained in dark brown and has an aromatic, bitter taste.

Table 1. Rendemen Results

Treatment	Weight sample	Weight Extract (g)	Rendemen (%)
5 minutes	50	13.9	27.8 %
10 minutes	50	12.2	24.4 %
15 minutes	50	12.1	24.2 %
20 minutes	50	14.3	28.6 %

Data source: Primary Data, 2019

### 3.3. Flavonoid qualitative test results

Qualitative flavonoids identification of kitolod leaves (*Isotoma longiflora* (L.) C. Presel) positively by showing a yellow formed, when extract reacted with powder Mg and HCl concentrated. This is in line with the study conducted by Hapsari Atika [3] where at the time of addition of reagents appeared yellow indicating the presence of flavonoid compounds. This is because flavonoids include phenolic compounds, when the phenol is treated with the base it forms a color due to the conjugation system of the aromatic group.

### 3.4. Maximum Wave Length and Absorption

Table 2. Maximum Wave Length and Absorption

wavelength (nm)	Absorbance
431	0.7450

Data source: Primary Data, 2019

Setting the maximum wavelength for measurement at the maximum absorption wavelength will result in maximum absorption, at the maximum wavelength having maximum sensitivity due to the greatest absorbance change. The resultant maximum wavelength of the standard solution was 431.0 nm with an absorbance of 0.7450.

### 3.5. Operating Time (OT) setting

Operating time was performed with the aim of knowing the stable measurement time when the sample reacted perfectly to the color reagent, the stabilization time being 45 minutes.

### 3.6. Standard Curve Making

The results of the concentration and absorbance of the standard curve solution can be seen in table 3:

Table 3. Concentration of Standard Curve Solution

Concentration (ppm)	Absorbance
10	0.259
20	0.330
30	0.403
40	0.485
50	0.562

Data source: 2019 Primary Data

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The concentration variation is used to detect differences in absorbance, where the higher the concentration of absorbance values the greater.

The production of a standard curve aims to obtain the standard solution of equilibrium in the determination of the sample rate, the relationship between the quercetin concentration and the absorption, the data obtained a correlation coefficient (r) of 0.9993 approaching 1 shows that the regression equation is linear.

Based on table 3 we obtain  $y = 0.0076x + 0.1795$  with values of  $r = 0.9993$  indicating that these values are linear.

### 3.7. Determination of Flavonoid Level

Determination of total flavonoid readings with absorbance at a maximum wavelength of 431 nm was obtained with total flavonoid content of kitolod leaves (*Isotoma longiflora* (L.) C. Presl) of 5 min incubation at an average rate of 0.31995% w/w or 31.9 mg / kg, 10 min infusion 0.25635% w / w or 25.6 mg / kg, 15 min 0.2239% w/w or 22.3 mg / kg, 20 min 0.17254% w/w or 17.2 mg / kg . Based on the observational data obtained test statistically using the One-Kolmogorov-Smirnov Test obtained  $0.992 > 0.05$  ( $H_0$  accepted). Then, with the one-way Anova test, homogeneity was found to be  $0.504 > 0.05$  ( $H_0$  accepted). From the Anova test data show that the sign of  $0.000 < 0.05$  then  $H_0$  is rejected, followed by LSD test. From the data between treatments it can be said that there is a significant difference between the treatments that is in each comparison of the time of reduction of flavonoid levels.

## 4. Conclusion

The longer the boiling time affects the total flavonoid content of the leaves kitolod, (*Isotoma longiflora* (L.) C. Presl), the longer the decomposition of the flavonoid content compound decreases.

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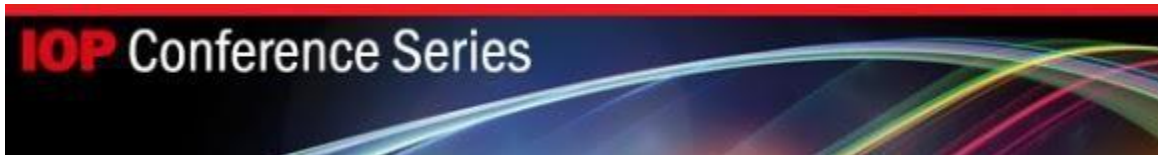
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Muchson Arroseyid<sup>1\*</sup>, Choiril Hana Mustofa<sup>1</sup>, Sutaryono<sup>1</sup>, and Alifiasri Praptiwi Rohmah<sup>1</sup>

<sup>1</sup> Department of Pharmacy, School of Health Science Muhammadiyah Klaten, Central Java

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## 3. Results and Discussion

### 3.1. Determined plant

The results of the plant determination indicate that this plant belongs to the family Lobeliaceae, a species (*Isotoma longiflora* (L.) C. Presel)

### 3.2. Extract Results

The extraction results of kitolod leaves (*Isotoma longiflora* (L.) C. Presel) were performed by varying the 5-minute, 10-minute, 15-minute, and 20-minute boiling method with aquadest solvent. 50 grams of dried kitolod leaves were obtained from 13.9 grams, 12.2 grams, 12.1 grams, 14.3 grams. The extract of kitolod leaves (*Isotoma longiflora* (L.) C. Presel) obtained in dark brown and has an aromatic, bitter taste.

**Table 1.** Rendemen Results

Treatment	Weight sample	Weight Extract (g)	Rendemen (%)
5 minutes	50	13.9	27.8 %
10 minutes	50	12.2	24.4 %
15 minutes	50	12.1	24.2 %
20 minutes	50	14.3	28.6 %

Data source: Primary Data, 2019

### 3.3. Flavonoid qualitative test results

Qualitative flavonoids identification of kitolod leaves (*Isotoma longiflora* (L.) C. Presel) positively by showing a yellow formed, when extract reacted with powder Mg and HCl concentrated. This is in line with the study conducted by Hapsari Atika [3] where at the time of addition of reagents appeared yellow indicating the presence of flavonoid compounds. This is because flavonoids include phenolic compounds, when the phenol is treated with the base it forms a color due to the conjugation system of the aromatic group.

### 3.4. Maximum Wave Length and Absorption

**Table 2.** Maximum Wave Length and Absorption

wavelength (nm)	Absorbance
431	0.7450

Data source: Primary Data, 2019

Setting the maximum wavelength for measurement at the maximum absorption wavelength will result in maximum absorption, at the maximum wavelength having maximum sensitivity due to the greatest absorbance change. The resultant maximum wavelength of the standard solution was 431.0 nm with an absorbance of 0.7450.

### 3.5. Operating Time (OT) setting

Operating time was performed with the aim of knowing the stable measurement time when the sample reacted perfectly to the color reagent, the stabilization time being 45 minutes.

### 3.6. Standard Curve Making

The results of the concentration and absorbance of the standard curve solution can be seen in table 3:

**Table 3.** Concentration of Standard Curve Solution

Concentration (ppm)	Absorbance
10	0.259
20	0.330
30	0.403
40	0.485
50	0.562

Data source: 2019 Primary Data



The concentration variation is used to detect differences in absorbance, where the higher the concentration of absorbance values the greater.

The production of a standard curve aims to obtain the standard solution of equilibrium in the determination of the sample rate, the relationship between the quercetin concentration and the absorption, the data obtained a correlation coefficient ( $r$ ) of 0.9993 approaching 1 shows that the regression equation is linear.

Based on table 3 we obtain  $y = 0.0076x + 0.1795$  with values of  $r = 0.9993$  indicating that these values are linear.

#### 2.8. Determination of Flavonoid Level

Determination of total flavonoid readings with absorbance at a maximum wavelength of 431 nm was obtained with total flavonoid content of kitolod leaves (*Isotoma longiflora* (L.) C. Presl) of 5 min incubation at an average rate of 0.31995% w/w or 31.9 mg / kg, 10 min infusion 0.25635% w / w or 25.6 mg / kg, 15 min 0.2239% w/w or 22.3 mg / kg, 20 min 0.17254% w/w or 17.2 mg / kg . Based on the observational data obtained test statistically using the One-Kolmogorov-Smirnov Test obtained  $0.992 > 0.05$  ( $H_0$  accepted). Then, with the one-way Anova test, homogeneity was found to be  $0.504 > 0.05$  ( $H_0$  accepted). From the Anova test data show that the sign of  $0.000 < 0.05$  then  $H_0$  is rejected, followed by LSD test. From the data between treatments it can be said that there is a significant difference between the treatments that is in each comparison of the time of reduction of flavonoid levels.

## 4. Conclusion

The longer the boiling time affects the total flavonoid content of the leaves kitolod (*Isotoma longiflora* (L.) C. Presl), the longer the decomposition of the flavonoid content compound decreases.

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